# Structure, function and evolution of the Crimean Congo hemorrhagic fever nucleocapsid protein

Stephen Carter, Cheryl Walter, Rebecca Surtees, Antonio Ariza, John Barr and Thomas Edwards

# Introduction

Recent research in our lab has focused on the Crimean-Congo hemorrhagic fever virus (CCHFV). CCHFV is a member of the *Bunyaviridae* family, and together with members of the *Arenaviridae* and *Orthomyxoviridae* families these viruses are known as segmented negative stranded RNA viruses (sNSV) by virtue of their multistranded genomes.

While these viruses are extremely diverse in their disease causing ability, they possess one common structural characteristic that is at the core of their respective life cycles; a ribonucleocapsid assembly (RNP). This is an association of the RNA genome with a virus-encoded nucleocapsid (N) protein, and its formation is essential for several fundamental aspects of the virus replication cycle including gene expression and virus assembly.

One aspect of our research is to try to understand how the structure of the CCHFV N RNP protein dictates and relates to its function. Towards this aim, we have solved the crystal structure of the CCHFV N protein to 2.1 Å. Our work describes the N protein structure, and the high degree of structural homology between the CCHFV N protein and the N protein from another sNSV member Lassa virus (LASV), which is an arenavirus. Furthermore the crystal structure of the CCHFV N protein guided site-directed mutagenesis of specific N protein residues, which delineate how N binds RNA and thus forms a competent RNP template for both RNA replication and mRNA transcription.

### Results

The CCHFV N is composed of a globular core comprising 15 alpha helices ( $\alpha$ 1-8;  $\alpha$ 14-20) and a prominent additional structural element we termed the 'arm' comprising two long alpha helices ( $\alpha$ 9-10) extending away from the core, presenting an exposed loop at its apex, and supported by a small three helix bundle ( $\alpha$ 11-13; Figure 1A). Electrostatic surface potential suggests a possible RNA binding 'platform' adjacent to the arm. Additional results from CCHFV mini-replicon system show some of these residues are important in CCHFV-specific RNA synthesis.



Structural comparisons indicate that the CCHFV N globular domain exhibits a high degree of structural homology with the N-terminal domain of LASV N, a member of the *Arenaviridae* 

family, whilst essentially structurally un-related to Rift Valley fever virus (RVFV), a member of the *Bunyaviridae* family. This data suggests the taxonony of two and three segmented RNA viruses may need re-examining.

The structural alignment provided additional clues about the RNA binding mechanism. The helix equivalent to LASV N  $\alpha$ 5 (yellow, Figure 2) is the helix in CCHFV N,  $\alpha$ 11, which precedes the long flexible loop at the base of the arm structure. This may represent a strong candidate for providing a similar RNA gating mechanism to that proposed for LASV N; RNA gating is mediated by a movement of  $\alpha$ 5 in the LASV N. Electrostatic surface of CCHFV N revealed an additional RNA binding 'pocket' equivalent to the LASV N RNA binding surface.



# **Publications**

Carter, S., Barr, J. & Edwards, T. (2012) Expression, purification and crystallization of the crimean-congo haemorrhagic fever virus nucleocapsid protein. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.* **68**: 569-573.

Carter, S., Surtees, R., Walter, C., Ariza, A., Bergeron, E., Nichol, S., Hiscox, J., Edwards, T. & Barr, J. (2012) Structure, function, and evolution of the crimean-congo hemorrhagic fever virus nucleocapsid protein. *J. Virol.* **86**: 10914-10923.

### Collaborators

*External:* Éric Bergeron and Stuart Nichol (Viral Special Pathogens Branch, CDC, Atlanta, USA).